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**Making teeth to order: conserved genes reveal an ancient molecular pattern in
paddlefish (Actinopterygii)**

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Summary

Ray-finned fishes (Actinopterygii) are the dominant vertebrate group today (+30,000
species, predominantly teleosts), with great morphological diversity, including their
dentitions. How dental morphological variation evolved is best addressed by
considering a range of taxa across actinopterygian phylogeny; here we examine the
dentition of *Polyodon spathula* (American paddlefish), assigned to the basal group
Acipenseriformes. Although teeth are present and functional in young individuals of
Polyodon, they are completely absent in adults. Our current understanding of
developmental genes operating in the dentition is primarily restricted to teleosts; we

show that *shh* and *bmp4*, as highly conserved epithelial and mesenchymal genes for gnathostome tooth development, are similarly expressed at *Polyodon* tooth loci, thus extending this conserved developmental pattern within the Actinopterygii. These genes map spatio-temporal tooth initiation in *Polyodon* larvae and provide new data in both oral and pharyngeal tooth sites. Variation in cellular intensity of *shh* maps timing of tooth morphogenesis, revealing a second odontogenic wave as alternate sites within tooth rows, a dental pattern also present in more derived actinopterygians. Developmental timing for each tooth field in *Polyodon* follows a gradient, from rostral to caudal and ventral to dorsal, repeated during subsequent loss of teeth. The transitory *Polyodon* dentition is modified by cessation of tooth addition, and loss. As such, *Polyodon* represents a basal actinopterygian model for the evolution of developmental novelty: initial conservation, followed by tooth loss, accommodating the adult trophic modification to filter-feeding.

Keywords

Polyodon, teeth, dentition, *shh*, *bmp4*, Acipenseriformes, paddlefish, evolution

1. Introduction

Most tooth development models reflect a bias toward morphologically derived vertebrates (e.g., zebrafish, mouse). However, more representative models for the evolution of developmental mechanisms of the dentition are provided by taxa at the base of extant vertebrate phylogenies. The basal actinopterygian Order Acipenseriformes includes fossil taxa as well as the American paddlefish *Polyodon* (Family Polyodontidae) and sturgeons (Family Acipenseridae, e.g. *Acipenser* [1, 2]) and represents an increasingly utilized system for addressing developmental questions in an

51 evolutionary context [3-6]. Due to their basal phylogenetic position, Acipenseriformes
52 are a particularly relevant model to test hypotheses of tooth patterning and evolution.
53 The dentition is lost in adult paddlefish and sturgeon, but present in younger
54 individuals, although details of early stages of tooth development are poorly known [1-
55 3, 7]. As pattern order for the forming dentition has previously been described for more
56 derived actinopterygians, comparable data for *Polyodon* will provide significant
57 information on mechanisms in more phylogenetically basal actinopterygians.

58 The secreted protein sonic hedgehog (*shh*) and the TGF- β superfamily member
59 bone morphogenetic protein4 (*bmp4*) are key dental patterning genes in vertebrates. *In*
60 *situ* hybridisation assays demonstrate that the transcripts coding for *shh/bmp4* are
61 present at the earliest sites of tooth initiation with focused, time specific loci of
62 expression restricted to dental epithelium (*shh*) [8, 9] and co-expression in the
63 underlying condensed mesenchyme (*bmp4*). Co-expression occurs on each
64 oropharyngeal dentate field, from a diffuse band of dental competence (odontogenic
65 band), to discrete placodes of single tooth initiation. Non-mammalian vertebrates for
66 which this conserved pattern of *shh/bmp4* expression has been used to characterize
67 dental patterning include a variety of teleosts (Osteichthyes, Actinopterygii): rainbow
68 trout (*Onchorhynchus mykiss* [8, 9]), Mexican tetra (*Astyanax mexicanus* [10]),
69 zebrafish (*Danio rerio* [11, 12]), several Lake Malawi cichlids [13], the freshwater
70 pufferfish (*Monotreta abei* [14]), as well the Queensland lungfish (*Neoceratodus*
71 *forsteri* [15]) and various snakes and lizards (Osteichthyes, Sarcopterygii, [16-18]) and
72 the catshark *Scyliorhinus canicula* [19, 20]. As the only non-teleost actinopterygian yet
73 surveyed, our new data from *Polyodon* will provide key phylogenetic support for the
74 hypothesis that *shh* and *bmp4* are part of a conserved and ancient gene regulatory
75 network for patterning vertebrate dentitions.

We predict that *Polyodon* will exhibit the conserved pattern of epithelial *shh*-positive loci, with comparable mesenchymal expression of *bmp4* [8], observed in other vertebrate taxa. Here we will use expression patterns for these genes, along with other histological and morphological datasets to demonstrate temporal differences in focal localization for each tooth site in *Polyodon*, mapping position and timing of tooth initiation to demonstrate how pattern order is established through co-ordinated gene activity. Our hypothesis is that this represents a basal condition of shared genetic regulation of tooth initiation times and topographic order for the Actinopterygii.

2. Material and Methods

(a) Animal care and sacrifice

Fertilized *Polyodon spathula* eggs were obtained from Osage Catfisheries, Inc. (Osage Beach, MO, USA) and raised to desired stages in recirculating, closed freshwater systems mimicking natural conditions (22°C, pH 7.2 ± 0.7, salinity of 1.0 ± 0.2 p.p.t. [21]). *Polyodon* were euthanized in a lethal dose of MS-222 (Tricaine) and fixed for at least 24h (dependent of tissue volume) in 4% paraformaldehyde [21]. All animal care, feeding, and euthanization protocols were in accordance with an approved IACUC (Institutional Animal Care and Use Committee) Animal Care Protocol [KSU #12-001; NSF IOS 1144965].

(b) Staging of larval *Polyodon*

Polyodon staging follows [3, 21]: lengths for individual specimens for stages 37-46, and other details of the staging, can be obtained from these. Feeding larvae (beyond stage 46) are described as ‘days post-staging’ (dps) and juveniles by standard length (SL). At incubation temperature (22°C), the larval period between hatching (stage 36) and onset of exogenous feeding and yolk exhaustion (stage 46) proceeds at approximately one

stage per 24 hour period [21].

(c) *in situ* hybridisation

In situ hybridisation used standard protocols [5] with riboprobes for *shh* [22] or *bmp4*. *Bmp4* was cloned from cDNA using the forward primer CGA GGC TAC TTT GTT GCA CA and reverse primer TCC ACG TAC AGT TCG TGT CG. Selected whole larvae (stages 41-45) with *shh* or *bmp4* expression were embedded in 20% gelatin and vibratome-sectioned at 50µm or, embedded in 30% sucrose, frozen in liquid nitrogen, and cryostat sectioned at 20µm. Numbers of specimens (antisense, comparable number of sense), *bmp* stages 34-39 n=7; 40-43 n=6; 44-46 n=6. *shh* stages 36 n=2; 38 n=3; 39 n=3; 40 n=2; 41 n=4; 42 n=2; 43 n=2; 45=6. Photomicrographs were taken with Zeiss Nomarsky optics, or an Olympus SZX16 dissecting microscope equipped with a QImaging RetigaEXi digital camera.

(d) Clearing and staining, CT-imaging

Cleared and stained specimens (CS; Alizarin red and Alcian Blue [23]) were dissected and mounted as half-jaws. Older specimens were studied as CS skeletal preps under a stereomicroscope and CT-scanned (X-Tek HMX ST CT scanner, Image and Analysis Centre, Natural History Museum, London; MicroCT at Dental Institute, King's College London, GE Locus SP, creating volumes with voxel sizes 6.5µm) and rendered using the software program Drishti (<http://sf.anu.edu.au/Vizlab/drishti>).

(e) Terminology

The terms distal and proximal are used in the upper and lower jaws, with reference to the jaw joint (proximal) and symphysis (distal). The terms rostral and caudal, dorsal and ventral are used with respect to the body axes.

3. Results

In *Polyodon spathula* larvae, *shh* and *bmp4* expression reveal both the early events of

oral and pharyngeal dental patterning, and sequential addition of tooth loci as development proceeds. There are noticeable differences in the addition of new tooth germs in individual dentate fields, normally caudal, but exceptionally rostrally on the palatopterygoid tooth plate. Concerning timing along the body axis, tooth initiation begins in association with Meckel's cartilage, establishing a spatio-temporal gradient that extends from the oral, through to tooth sites in the pharyngeal cavities (figures 1, 2; electronic supplementary material, figure 4). Skeletal preparations provide additional data on pattern order; after tooth rows form on the dentary and dermopalatine, they develop on the more caudal palatopterygoids and first hypobranchials (figures 1*a*, *c*, 2*a*, *b*, respectively). Teeth are later organized into toothed plates, connected by basal bone of attachment, representing functional surfaces of the oropharyngeal dentition (electronic supplementary material, figure 2*c*; Table) [1-3].

(a) Timing of *shh* expression in whole mounts maps sequential tooth initiation (stages 37-43)

Spatial expression of *shh* occurs as focal loci, with changes in intensity coincident with each stage of tooth germ morphogenesis, mapping location and developmental timing for each tooth position (figures 1, 3, 4). This pattern of spatio-temporal expression identifies new tooth germs added relative to preexisting ones, in precise locations at sequential times, from one dentate region to another (electronic supplementary material, Table 1).

Shh expression is first observed in the odontogenic fields beginning at stage 37 (figure 1*d*, electronic supplementary material, figure 4*b*). Strong expression loci on the odontogenic band occur first as focused placodes (stages 39-41; figures 1*a-e*), then expression as a cap around the cone of the tooth tip (figures 3*p*², 4, electronic

supplementary material, figure 4*c-h*). These loci mark tooth positions within one row (figure 1; electronic supplementary material, figure 4*i-p*). *Shh* expression is next upregulated at alternate (second) tooth positions, within this same row (figure 1*a-c*, *e*, arrows). By stage 43 *shh* is downregulated in epithelial cells of older tooth germs around tooth cones. Accurate counts of tooth number from *shh* expression at these later stages relies on seeing tooth cones (using Nomarsky optics). Nevertheless, differences in total number between upper and lower jaws are observed (electronic supplementary material, Table 1); for example, at stages 40 and 42 there are more tooth loci on the dentary than dermopalatine (compare electronic supplementary material, figure 4*g*, *h* (new parasymphysial tooth on dentary) with 4*e*, *f*, and 4*i-m* (new loci added distally) with 4*n*).

Given this recognizable developmental sequence of epithelial *shh* expression, sites of tooth initiation can be identified along the rostrocaudal body axis. In both jaws at stages 39-40, there are four to five tooth buds in each dentary and dermopalatine field, contrasting with lack of tooth buds in more caudal toothed sites (electronic supplementary material, figure 4*c-h*). Later, at stage 41 the dentary and dermopalatine have seven tooth positions with alternating higher intensity of *shh* expression, and a new distal and proximal tooth germ, all in the same tooth row (figure 1*e*, *i*, arrows). As well, two *shh*-positive tooth loci are present on the first hypobranchial and the palatopterygoids (figure 1*a-c*, *e*, arrowheads). In stages 42-43 these *shh* expression sites are intense caps around the tooth cone (electronic supplementary material, figure 4*la*, *lb*), forming rings in later stages where *shh* is downregulated in cap cells (electronic supplementary material, figure 4*o-p*, further details see §3*c*, *d*, and figure 4).

(b) *Bmp4* expression maps timing of co-operative activity during tooth

morphogenesis (stages 40-45, 1dps)

All stages show *bmp4* expression associated with each tooth locus (figure 1*f-i*; electronic supplementary material, figure 5). When compared to stage-matched specimens stained for *shh*, intense expression of *bmp4* appears associated with mesenchyme of the newest forming tooth loci (figure 1*g, i*, arrows). Notably, stage 41 and 45 *bmp4* expression shows upregulation in alternate positions of (second) tooth germs within the tooth row, on the dentary and dermopalatine, while the most rostral (first) tooth germs are dentine cones with *bmp4* downregulated in the papilla. Note these show strong papillary expression in more caudal sites, indicating that these are younger (figure 1*f, g, i*; asterisk versus arrows, respectively). However, on the palatopterygoid, the intense papillary *bmp4* expression of the younger loci is rostral to the dentine cones, as observed in the expression pattern for *shh* (i.e. an opposite second tooth addition pattern to the dentary and dermopalatine, electronic supplementary material, figure 5*h*’, st 42, 5*l*’, st 45, arrows

(c) Cellular expression of *shh* during tooth germ morphogenesis, stage 45

The exact location of expression within the epithelial tooth germ is shown in more detail in serial, parasagittal sections than in whole mount *in situs* (figure 3, electronic supplementary material, figure 4), while the mesenchyme of the dental papilla shows complimentary *bmp4* expression (electronic supplementary material, figure 6). Gene expression changes are associated with different tooth germ morphologies through development (figures 3*p, 4*), where different intensities are associated with specific timing of morphogenesis at each tooth site in the oropharyngeal cavity, including first locations of the sites on the branchial arches. These demonstrate a rostro-caudal activation gradient of tooth initiation for each dentate field. Initially, the placode shows intense *shh* and *bmp4* expression and is superficial (no dental lamina), with *shh* located

to the middle epithelial cells (figure 3*d, i, p*¹). In the cap stage, *shh* is more intense in all epithelia, surrounding the papilla (figure 3*g, p*²; *bmp4*, electronic supplementary material, figure 6*b*). After dentine histogenesis, *shh* is downregulated in the cap cells but is strongly expressed in the epithelium as a collar around the tooth cone (cone+collar stage, figure 3*c, p*³). Subsequently, *shh* is downregulated around the whole tooth cone (figure 3*j, n, p*⁴), but within the adjacent dental epithelium (not the inner dental epithelium), *shh* is upregulated as an intense focal expression, attributed to an incipient, successive tooth germ (figures 3*j, n*; electronic supplementary material, figure 6*a, c, d*). In the second, alternate tooth position the same steps of *shh* expression are observed, including cap, and cone+collar stages (figure 3*f, h, k*).

Serial sections show these expression stages simultaneously throughout the oropharyngeal cavity. Loci of *shh* expression occur dorsally on the dermopalatine and palatopterygoid (figure 3*a-d, g*), and ventrally on the dentary and 1st hypobranchial (figure 3*e, h, i*; electronic supplementary material, figure 6*a*), along with a focal spot on the infrapharyngobranchials dorsally and 1st and 2nd hypobranchials ventrally (figure 3*e, h, i*; electronic supplementary material, figure 6*a, d*), but a field of expression on the more caudal branchial arches (figure 3*e*). When dentine is present in the first dentary teeth, as a collar plus translucent cone, the more caudal, second tooth germ is only at the placode stage (figure 3*f*). In other sections, the first tooth appears as a translucent dentine cone with a second tooth at cap, or collar stage (figure 3*h*). All these observations show a staggered time difference in each second tooth germ, as well as the first (*bmp4* data, electronic supplementary material, figure 6*c, d*). Similar staggered stages are seen in the dermopalatine tooth germs, and those of the palatopterygoid relative to the dermopalatine (figure 3*m, n, g*).

The restriction of *shh* expression to an intense focal locus (placode) forms first in the evaginated epithelium above the cartilage on the 2nd, as in the 1st, hypobranchial

(figure 3*l*). The placode is superficial (i.e. forms without a dental lamina; figure 3*n*, *o*, 3*p*¹), but also evaginated at the cone-cap stages (figure 3*h*, 3*p*²), then just within the expanded dental epithelium at cone+collar stage (figure 3*k*, *n*, 3*p*³). When *shh* is downregulated in all dental epithelium around the tooth there is an upregulated intense locus of *shh* expression next to this first tooth, in the dental epithelium, ‘cone+bud’, not evaginated but located in the epithelium adjacent to the dentine cone. Papillae with taste buds on the inner oral epithelium always exhibit faint *shh* expression, similar in intensity to the downregulated collar epithelium (figures 3*o*, arrow, 4*c*, sensory papilla with differentiated cells), while *bmp4* expression is absent (electronic supplementary material, figure 6).

(d) Skeletal preparations show tooth addition positions in 7dps larvae

i) Tooth development on upper jaw, dorsal branchial skeleton. Tooth rows are present ventral to the upper jaw cartilage, both rostrally on the dermopalatine bone and caudally on the palatopterygoid. The dermopalatine has 17-20 ankylosed teeth, while the latter lacks an independent ossification at this stage, with teeth conjoined by the individual bone of attachment of each tooth (translucent rings, figure 2*a*, *c*, *g*, *h*). Caudal to the palatopterygoid are two paired patches of teeth, the first associated with the hyoid arch with six teeth, joined only by their bases (figure 2*c*, black arrow, white box, *k*). The second is associated with the 2nd infrapharyngobranchial, possessing four teeth (figure 2*c*, white box, *j*). The dermopalatine bone represents the most developmentally advanced in the upper jaw with new unattached teeth being added caudal to tooth positions 2 and 4, as well as parasymphysially (arrows, figure 2*g*, *i*). On the ventral surface of the palatopterygoid cartilage the oldest teeth are joined together via attachment bone (dentine cones expanded into cylinders), with 11 teeth on the right

side, nine on the left. As opposed to the caudal tooth addition associated with the dermopalatine, two new teeth (lacking bony rings; figure 1g, *h*, arrows) are rostral to the attached (older) teeth.

ii) Tooth development on lower jaw, ventral branchial skeleton. Tooth rows are present dorsally on Meckel's cartilage, with 22 left and 21 right teeth fused to the dentary bone via bone of attachment with new, unattached teeth caudal to the attached (older) teeth and at proximal and distal ends of the row (Mc, figure 2d, *e*, arrows). Other toothed plates are caudal to Meckel's cartilage in the pharyngeal cavity, on the hypobranchials (first, 11 teeth; second, three teeth). Hypobranchial teeth are not ankylosed to bone but older teeth are joined at their bases via their individual bone of attachment (figure 2d, *f*, *l*). Three new teeth (not joined by bone of attachment) on left hypobranchial 1 are added caudally (figure 1f, arrows). By later functional stages, with increasing tooth numbers at all sites, pharyngeal teeth are arranged in radial rows (four to five teeth in each), differing from the oral dentition (electronic supplementary material, figure 2a, *b*).

Discussion

Combined data from ontogenetic stages of *Polyodon spathula* establishes sequences of gene expression and tooth morphogenesis in the oropharyngeal cavity, allowing spatiotemporal patterns of tooth initiation and development to be documented; tooth rows form on the dentary and dermopalatine before the more caudal palatopterygoids and first hypobranchials (figures 1a, *c*, 2a, *b*, respectively). Teeth are later organized into toothed plates, connected together by basal bone of attachment, independently of the membrane bone, representing early functional surfaces of the oropharyngeal dentition (electronic supplementary material, figure 2c). Skeletal whole mounts show where new teeth are added to individual dentate fields, while post-larval stages indicate that tooth

addition slows and teeth are lost (electronic supplementary material).

These observations indicate progressive rostral-caudal and ventro-dorsal tooth initiation/addition gradients within the oropharyngeal cavity: tooth addition occurs first on Meckel's cartilage, showing alternate patterns of gene expression along the tooth row, prior to the dermopalatine (stage 40, electronic supplementary material, figure 4g, *h*; stage 42, electronic supplementary material, figure 4*i-m* versus 4*n*). At 7dps, a larger number of teeth are present on the dentary (figure 2) and at later juvenile stages the dentary shows substantial toothless areas of membrane bone relative to other dentate regions in the oropharyngeal cavity, due to tooth related loss of attachment bone (electronic supplementary material, figures 1*f-i*, 3*c-f*, asterisk). With respect to a rostral-caudal gradient of tooth addition, the dentary and dermopalatine develop tooth germs with a cone of dentine before the palatopterygoid (electronic supplementary material, figure 5), while teeth in the oral cavity develop before those in the pharyngeal cavity. There is also a rostral-caudal progression in the pharyngeal cavity with the placode stage attained in hypobranchial 1, versus field expression on hypobranchials 2. The former has the most teeth; caudally hypobranchials 3 and 4 never show upregulated tooth loci. With respect to rostro-caudal tooth addition on each oral site, new tooth buds are initiated caudally on the dermopalatine and the dentary (figure 1*i*, electronic supplementary material, figure 5*m, n*), but new teeth form rostrally on the palatopterygoid (figure 2g, *h*).

Our results show that *shh* and *bmp4* expression data during *Polyodon* tooth initiation follows the same spatio-temporal order observed in all other non-mammalian vertebrate species assayed to date [8, 9, 14-17, 24]; however, our observations on the ordered sequence of timing of tooth germ initiation in oral and pharyngeal tooth sets also reveal directed rostro-caudal and ventro-dorsal patterns. This graded progression

has not previously been reported for actinopterygians, nor for gnathostome oropharyngeal dentitions. Nevertheless, tooth patterning, at least with respect to tooth initiation and differentiation appears evolutionarily stable and highly conserved among gnathostomes. For example, no differences in collocation of *shh* and *bmp4* expression were detected between developing oral and pharyngeal teeth in *Polyodon*, comparable to a variety of other taxa. Along with the ordered tooth initiation sequence, this implies that tooth germs in all regions are equivalent and conserved modular vertebrate units.

We have demonstrated cellular partitioning for *shh* and *bmp4* expression and sequential stages of tooth germ morphogenesis from ‘placode’, ‘cap’, ‘cone+collar’ to ‘cone +bud’ (figure 4). This is based on expression intensity that changes in a characteristic sequence within the dental epithelium, for each developing tooth germ. Notably, a new locus for strong expression forms alongside the developed, functional tooth (‘cone+bud’). We interpret this as the incipient tooth germ representing what we term a successional tooth. This is distinct from superficial, initial tooth ‘placodes’ and is consistent with observations that in actinopterygian fish, successional teeth form from the older tooth and not from a dental lamina [8]. In some actinopterygian taxa (Cyprinidae, derived teleosts), functional and replacement teeth can be retained as a pair, particularly during larval stages, although the functional tooth is eventually lost with the replacement tooth moving into place [25]. In *Polyodon*, by comparison, the functional tooth is retained and not lost in response to the presence of the successional tooth; the latter should therefore not be considered a replacement tooth *per se*. Tooth loss occurs much later in *Polyodon* in what appears to be a general reduction and loss in the oropharyngeal cavity. This suggests that the more typical osteichthyan dentition pattern, with tooth replacement, never happens and is altered at this early ontogenetic stage.

Despite the enormous diversity, the presence of teeth organized into a functional dentition is a shared feature among jawed vertebrates, undoubtedly one reason for their evolutionary success, allowing a variety of feeding niches to be exploited. This diversity is underpinned by a high degree of developmental genetic conservation, particularly in early development, in taxa such as trout [7, 8], cichlids [10, 24] and the pufferfish [12]; these early patterns are also seen in sarcopterygian fish *Neoceratodus* [13] as well as the shark *Scyliorhinus* [17, 26]. This conservation is also present in the dentition of *Polyodon spathula*, with modifications early in development, including tooth retention and lack of replacement teeth. Tooth addition slows, while in *Acipenser*, teeth are lost, entirely linked to suction feeding adaptations [1, 2]. However, we currently lack information on candidate genes involved in tooth regeneration that may change, or be missing in *Polyodon* and *Acipenser* [7]; other basal taxa, such as *Polypterus*, show full dentitions with tooth replacement [27]. New analysis of genes directed towards key transitions from tooth initiation to replacement in *P. spathula* will offer insight into the evolution of tooth regeneration strategies and dental diversity. Modifications to the dentition that occur later in ontogeny, allow the diversity of vertebrate dentitions to be expressed [10], and are the precursor steps to the development of drastically different modes of feeding among the gnathostomes.

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Data Accessibility.

Drishti files for scans of *Polyodon spathula* are available at <http://chondrichthyes.myspecies.info/>.

References

1 Grande, L. and Bemis, W. E. 1991 Osteology and phylogenetic relationships of fossil and recent paddlefishes (Polyodontidae) with comments on the interrelationships of Acipenseriformes. *J. Vert. Paleo. Spec. Mem.* **1**, 1–121.

2 Bemis, W. E., Findeis, E. K., and Grande, L. 1997 An overview of Acipenseriformes. *Env. Biol. Fishes* **48**: 25–71.

3 Bemis, W. E. and Grande, L. 1992 Early development of the actinopterygian head. I. External development and staging of the paddlefish *Polyodon spathula*. *J. Morph.* **213**, 47–83.

4 Davis M. C. 2013 The deep homology of the autopod: Insights from Hox gene regulation. *Int. Comp. Biol.* **53**, 224–234. (DOI: 10.1093/icb/ict029).

5 Modrell, M. S., Buckley, D. and Baker, C. V. H. 2011a Molecular analysis of neurogenic placode development in a basal ray-finned fish. *Genesis* **49**, 278–294.

6 Modrell, M. S., Bemis, W. E., Northcutt, R. G., Davis, M. C. and Baker, C. V. H. 2011b Electrosensory ampullary organs are derived from lateral line placodes in

- bony fishes. *Nature Comm.* **2** (496), 3142–3146. (DOI: 10.1038/ncomms1502).
- 7 Hilton, E. J., Grande, L. and Bemis, W. E. 2011 Skeletal anatomy of the shortnose
sturgeon, *Acipenser brevirostrum* Lesueur 181, and the systematics of sturgeons
(Acipenseriformes, Acipenseridae). *Fieldiana (Life Earth Sci.)* **3**, 1–168.
- 8 Fraser, G. J., Graham, A. and Smith, M. M. 2004 Conserved deployment of genes
during odontogenesis across osteichthyans. *Proc. Biol. Sci.* **271**, 2311–2317.
- 9 Fraser, G. J., Berkovitz, B. K., Graham, A. and Smith, M. M. 2006 Gene deployment
for tooth replacement in the rainbow trout (*Oncorhynchus mykiss*): a
developmental model for evolution of the osteichthyan dentition. *Evol. Dev.* **8**,
446–457.
- 10 Stock, D. W., Jackman, W. R. and Trapani, J. 2006 Developmental genetic
mechanisms of evolutionary tooth loss in cypriniform fishes. *Development*, **133**,
3127–3137.
- 11 Jackman, W. R., Yoo, J. J. and Stock, D. W. 2010. Hedgehog signaling is required at
multiple stages of zebrafish tooth development. *BMC Dev. Biol.* **10**, 119.
- 12 Wise, S. B. and Stock, D. W. 2006. Conservation and divergence of *Bmp2a*, *Bmp2b*, and
Bmp4 expression patterns within and between dentitions of teleost fishes. *Evol. Dev.* **8**,
511–523.
- 13 Fraser, G. J., Bloomquist, R. F. and Streelman, J. T. 2008 A periodic pattern
generator for dental diversity. *BMC Biol.* **6**, 32.
- 14 Fraser, G. J., Britz, R., Hall, A., Johanson, Z., and Smith, M. M. 2012 Replacing the
first-generation dentition in pufferfish with a unique beak. *Proc. Natl. Acad. Sci.*
USA **109**(21), 8179–8184.
- 15 Smith, M. M., Okabe, M. and Joss, J. 2009 Spatial and temporal pattern for the
dentition in the Australian lungfish revealed with sonic hedgehog expression

- 401 profile. *Proc. Biol. Sci.* **276**, 623–631.
- 402 16 Buchtova, M., Handrigan, G. R., Tucker, A. S., et al. 2008 Initiation and patterning
403 of the snake dentition are dependent on Sonic hedgehog signaling. *Dev. Biol.* **319**,
404 132–145.
- 405 17 Richman, J. M. and Handrigan, G. R. 2011 Reptilian tooth development. *Genesis* **49**,
406 247–260. (DOI: 10.1002/dvg.20721).
- 407 18 Handrigan, G. R. and Richman, J. M. 2010 Autocrine and paracrine Shh signaling
408 are necessary for tooth morphogenesis, but not tooth replacement in snakes and
409 lizards (Squamata). *Dev. Biol.* **337**, 171–186.
- 410 19 Smith, M. M., Fraser, G. J., Chaplin, N., Hobbs, C. and Graham, A. 2009 Reiterative
411 pattern of sonic hedgehog expression in the catshark dentition reveals a
412 phylogenetic template for jawed vertebrates. *Proc. Biol. Sci.* **276**, 1225–1233.
- 413 20 Fraser, G. J. and Smith, M. M. 2010 Evolution of development for patterning
414 vertebrate dentitions: an oro-pharyngeal specific mechanism. *J. Exp. Zool. B Mol.*
415 *Dev. Evol.* **314B**, 99–112 (DOI: 10.1002/jez.b.21387).
- 416 21 Davis, M. C., Shubin, N. H., and Force, A. 2004 Pectoral fin and girdle development in the
417 basal actinopterygians *Polyodon spathula* and *Acipenser transmontanus*. *J. Morph.* **262**,
418 608–628 (DOI: 10.1002/jmor.10264).
- 419 22 Davis, M. C., Dahn, R. D. and Shubin N. H. 2007 An autopodial-like pattern of Hox
420 expression in the fins of a basal actinoptergian fish. *Nature* **447**, 473–476.
- 421 23 Taylor, W. R. and van Dyke, G. C. 1985 Revised procedures for staining and
422 clearing small fishes and other vertebrates for bone and cartilage study. *Cybium* **9**,
423 107–119.
- 424 24 Fraser, G. J., Hulsey, C. D., Bloomquist, R. F., et al. 2009 An ancient gene network
425 is co-opted for teeth on old and new jaws. *PLoS Biol.* **7**, e31.

- 426 25 Van der Heyden, C. and Huysseune, A. 2000. Dynamics of tooth formation and
427 replacement in the zebrafish (*Danio rerio*) (Teleostei, Cyprinidae). *Dev. Dyn.* **219**,
428 486–496.
- 429 26 Tucker, A. S. and Fraser, G. J. 2014. Evolution and developmental diversity of tooth
430 regeneration. *Semin. Cell Dev. Biol.* 25–26, 71–80 (DOI:
431 10.1016/j.semcdb.2013.12.013).
- 432 27 Clemen, G., Bartsch, P. and Wacker, K. 1998 Dentition and dentigerous bones in
433 juveniles and adults of *Polypterus senegalus* (Cladistia, Actinopterygii). *Ann.*
434 *Anat.* **180**, 211–221.
- 435
- 436

Figure Captions

Figure 1. Expression of *shh*, *bmp4* in *Polyodon spathula* oral and pharyngeal initial

dentitions, stage 41. (*a-c*, *e*) *shh* expression in tooth buds of cleared whole mount jaws

compared with (*d*) stage 37 upper jaw, expression restricted to oral surfaces and on first

infrapharyngobranchial arches. (*a*, *c*) multiple loci on tooth fields of dentary and

dermopalatine, only two loci on hypobranchial and palatopterygoid. Arrows indicate

alternate timing of strongest expression. (*b*, *e*) strong expression in hypobranchial 1 and

palatopterygoid (arrowheads); cone expression in dentary, hypobranchial,

dermopalatine, compared to early placode expression on palatopterygoid. (*f-i*) *bmp4*

expression for comparison to *shh* expression. (*f*, *g*) lower jaw, (*h*, *i*) upper jaw *bmp4* in

the dental papillary mesenchyme marks all oral jaw tooth positions. Dental mesenchyme

underlies the dental epithelium and expression appears diffuse, however, more intense

expression is seen at alternate tooth loci (arrows, *f*, *g*, *i*) with weaker expression

indicating earlier (older) loci (asterisk), equivalent to *shh* expression pattern.

Abbreviations: b1, 2, basibranchials; ba, bone of attachment, cb1-5, ceratobranchials;

ch, ceratohyal; de, dentary; d.pal, dermopalatine; hb1, 2, 1st, 2nd hypobranchial; hb1tp,

hb2tp, hypobranchial toothplates; hh, hypohyal; hym, hyomandibular; itg, incipient

tooth germ; iph, infrapharyngobranchial; iphtp, infrapharyngobranchial toothplate; Mc,

Meckel's cartilage; ppt, palatopterygoid; tc, tooth cone; 2ndt, second tooth.

Figure 2. Alizarin red, Alcian blue preparations of *Polyodon spathula*, 7dps

showing relative tooth positions. (*a*, *c*, *g-k*) upper jaw and dorsal pharyngeal skeleton,

(*b*, *d-f*, *l*) lower jaw and ventral pharyngeal skeleton. (*a*, *b*) chondrocranium and

branchial arches. (*c*) upper jaw, teeth along dermopalatine bone and separate

palatopterygoid tooth plate (lacking membrane bone), with two paired tooth plates

caudally (black arrows indicate *j*, *k*). (*d*) lower jaw, ventral pharyngeal skeleton (hyoid, 1st, 2nd gill arches). (*e*) teeth on dentary bone (arrows, new teeth). (*f*) eight teeth linked by bone of attachment on 1st gill arch cartilage (hypobranchial 1), lacking membrane bone). (*g*-*i*) left upper jaw, teeth ankylosed to dermopalatine bone, separate palatopterygoid tooth plate (arrows, new teeth caudally on dermopalatine (*i*), rostrally on palatopterygoid (*h*)). (*h*) palatopterygoid tooth plate, bone of attachment only (arrows new teeth). (*j*, *k*) upper jaw tooth plates of (*j*) epibranchial 2, four associated teeth, (*k*) hyoid arch, six teeth. (*l*) hypohyal and first two ventral gill arches, with paired toothplates, more teeth on hb1 than hb2, more on ventral than dorsal pharyngeal toothplates. White arrows=newest unattached teeth. Scale bars *a*, *b*=1mm, *c*-*g*, *l*=500um, *h*, *i*=100um; abbreviations as in figure 1.

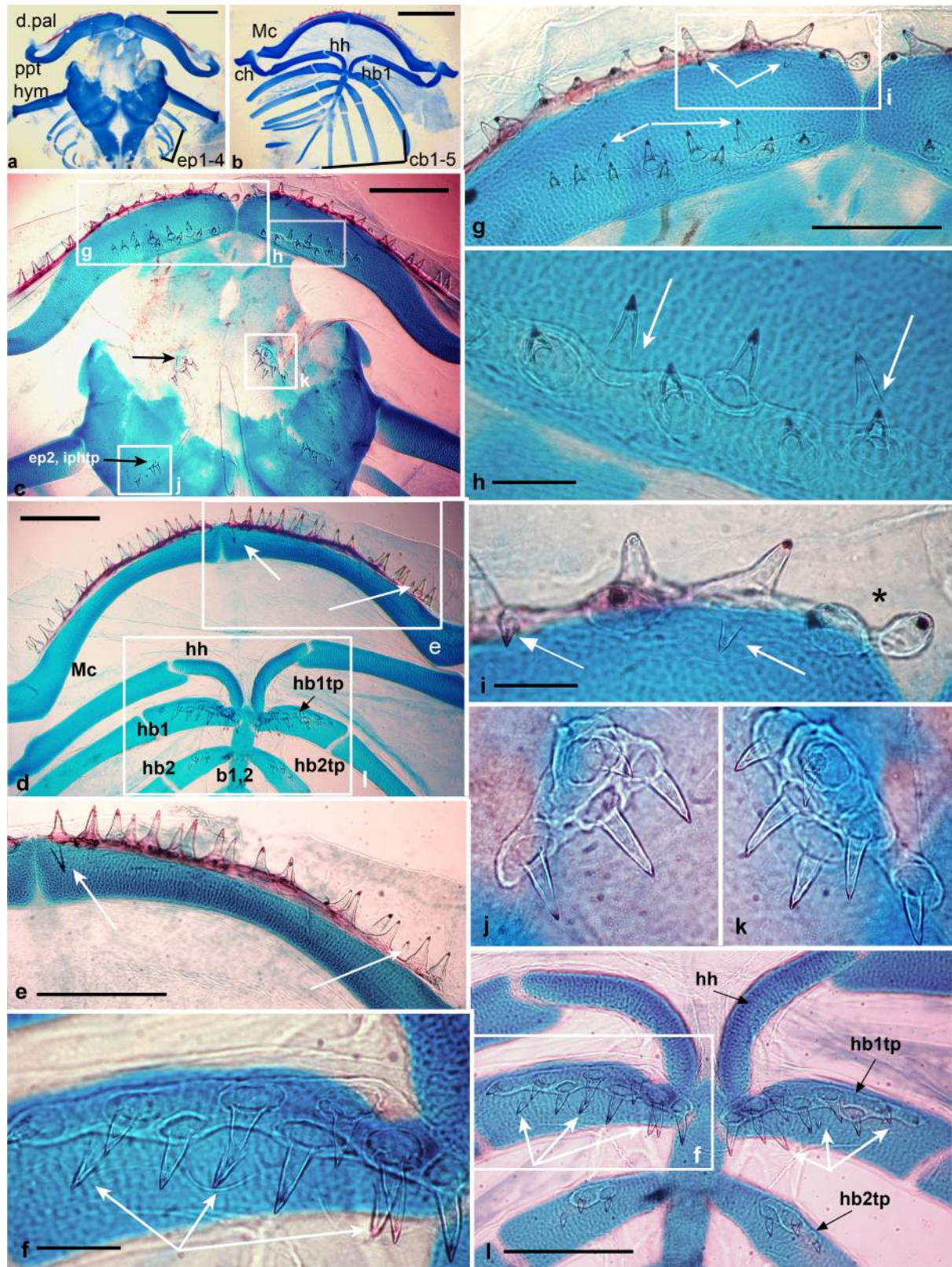
Figure 3. Serial sagittal sections, *Polyodon spathula* (stage 45) after *in situ* hybridisation for *shh* show sequence of tooth morphogenesis. Photomicrographs, low and high magnification (objectives x6.3, x16, x40) of location and rostro-caudal timing of *shh* gene expression in all tooth fields relative to tooth germ morphogenesis, rostral, left and dorsal, top. (*a-d*) most medial section, expression in dermopalatine (cone + collar, p^3) and palatopterygoid (placode, p^1). (*e*) more lateral section including Meckel's cartilage and pharyngeal arches. Expression loci associated with first stages of morphogenesis (placode, p^1) on the 1st upper branchial arch (iph1), 1st and 2nd hypobranchials. By comparison, on 3rd and 4th pharyngeal arches tooth buds foci absent, localisation is a field of expression, a stage prior to tooth morphogenesis. (*f*) low magnification field of variation in expression loci on dentary and hypobranchial1, with collar epithelium downregulated on first tooth (asterisk) and adjacent second tooth germ shown as intense expression (arrowhead, weak expression in sensory papilla, arrow as

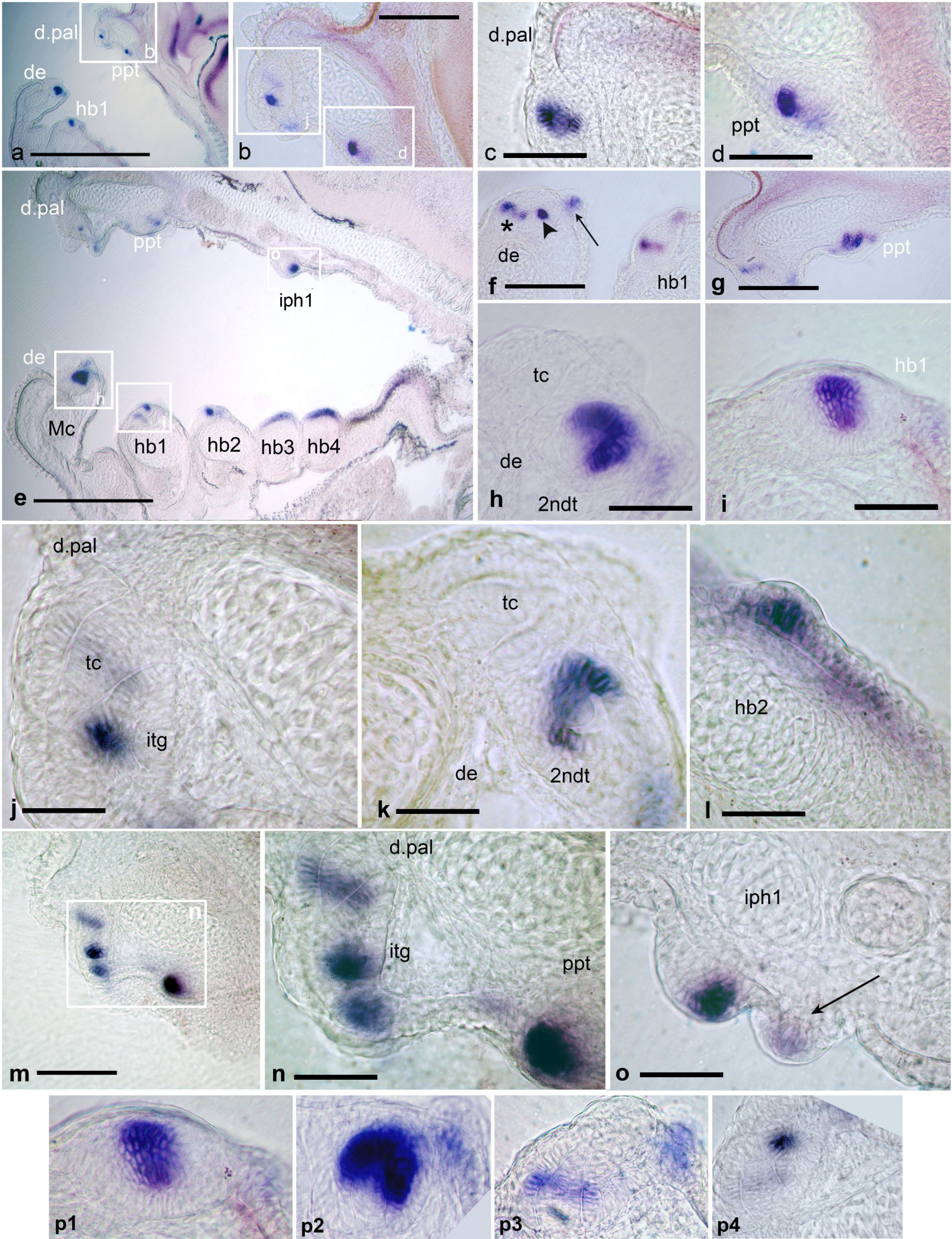
(*o*, *p4*). (*g*) low magnification view of variation in expression at loci on the dermopalatine (downregulated) and palatopterygoid strong expression in all dental epithelium around dentine cone (late cap stage). (*h*) tooth cone (tc) developed, and 2nd tooth germ (2ndt) at cap stage (*p*²). (*i*) 1st hypobranchial, placode stage of *shh* expression (*p*¹). (*j*) tooth cone with second incipient tooth germ (itg), strong expression (*p*⁴). (*k*) downregulation from cap to ‘collar’ expression (*p*³) in 2nd tooth. (*l*) early tooth placode in oral epithelium of 2nd hypobranchial. (*m*) upper jaw palatoquadrate cartilage with tooth germs on dermopalatine and palatopterygoid at different morphogenetic stages. (*n*) four stages of *shh* expression, tooth cone with downregulated expression, incipient second tooth germ on dermopalatine, on palatopterygoid, cap stage. (*o*), infrapharyngobranchial (iph1) upregulated strong expression (note evaginated tooth germ, placode-cap), alongside weak expression in sensory papilla (arrow). (*p*¹⁻⁴) four stages of *shh* expression in tooth germs, oral epithelium dorsal, contrast enhanced (translated into diagram as figure 4 a-d). Scale bars *a*, *e* = 250um, *b*, *f*, *g*, *m* = 50um, *c*, *d*, *h-l*, *n-p1-4* = 25um; abbreviations as in figure 1.

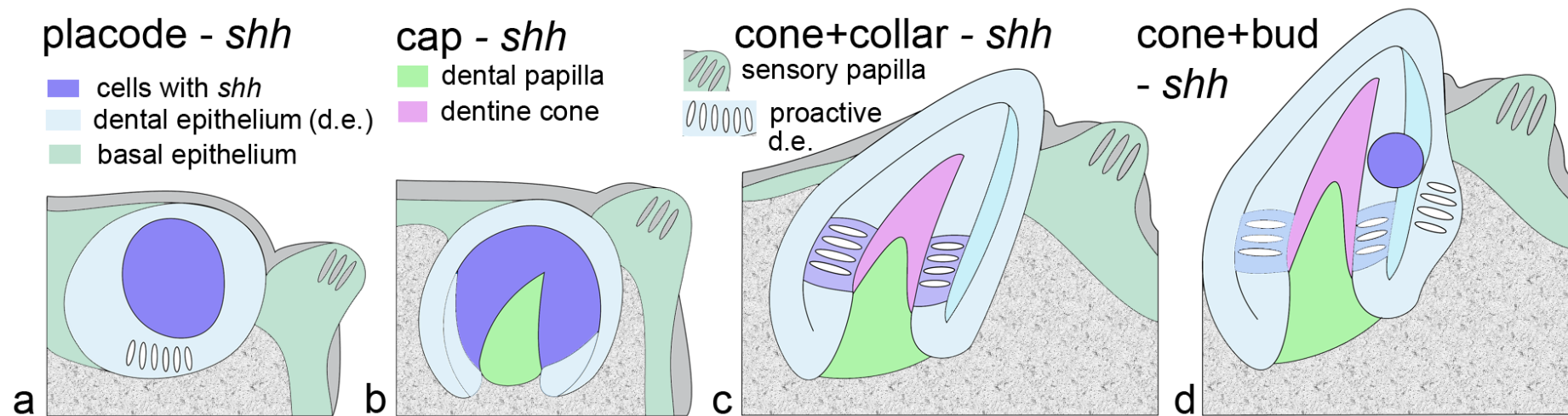
Figure 4. Diagram summarising stages of tooth germ morphogenesis relative to *shh* expression (from figure 3*p*¹⁻⁴). Intensity of cellular expression is partitioned characteristically within the dental epithelium, with negative differentiated, interactive cells of dental epithelium shown (proactive d.e.), and also in sensory papilla of taste buds on right of tooth germ. (*a*) cellular partitioning of *shh* expression as ‘placode’ (localized within epithelium, can be evaginated). (*b*) ‘cap’, expression in the cap-shaped epithelium of tooth germ, surrounding dental papilla. (*c*) ‘cone+collar’, cones of dentine with expression associated with the tooth base, or collar epithelium below the cap. (*d*) ‘cone +bud’, expression in a new site within the outer dental epithelia (incipient bud for

512 new tooth germ).









Specimen	UJ d.pal	UJ ppt	LJ de	UBS iph	UBS epb	LBS hb1	LBS hb2	Figure number
St. 39-40 <i>shh</i>	4+1	0	5	0	0	0	0	ESM 4c-h
St. 41-42 <i>shh</i>	4+2	1+1	7	0	0	1+1	0	1, ESM 4a, i-n
St. 43 <i>shh</i>	8+	2	11+	0	0	1	0	4o, p
Stage 45 <i>shh/bmp</i>	14	8	16-18	2	0	4	0-1	3, ESM 6
7dps larva *	17-20	9-11	21-22	4-6	4	11	3-4	2
TL-345 mm **	55	55	91	0	0	30	10	ESM 1, 2

Table 1: Rostro-caudal and ventro-dorsal graded trends from oral to pharyngeal sites in tooth addition during development and transition of the embryo to juvenile dentition, *Polyodon spathula*. Differences in total tooth number at each stage of development are shown and reflect a directed pattern in time and space throughout the oro-pharyngeal cavity. Abbreviations: de, dentary; d.pal, dermopalatine; epb, epibranchial; hb1, 2, hypobranchial 1, 2; iph, infrapharyngobranchial; ppt, pterygopalatine; UBS, LBS, upper, lower branchial skeleton; UJ, LJ, upper, lower jaw. *Based on n=5 cleared and stained whole mount. **Based on CT scan data. Numbers are per left or right half.